

# Remediation of Pesticide-Polluted Water using Ozonation as a Safe Method

Prof. Dr. Ahmed Khamis Salama<sup>1</sup>

<sup>1</sup> Majmaah University, KSA

*Received: 16 December 2012 Accepted: 31 December 2012 Published: 15 January 2013*

---

## Abstract

The effectiveness of ozone to remove the organic compound, chloropyrifos or cypermethrin from water at the two different levels of 1 and 2 ppm, for each and different contact times was studied. The recovered amounts of chloropyrifos or cypermethrin were extracted based on the solid phase extraction (SPE) method and then analyzed by GC-MS. The results demonstrated that the removal of these organic compounds by ozone increased with increasing the contact time. The removal percentages of chloropyrifos following ozone bubbling for different periods of 15, 30, 45 and 60 minutes at room temperature were 10.5, 96.3, 97.4 and 98.5

---

## Index terms—

## 1 Introduction

he excessive use of pesticides, their volatility and long-distance transports eventually results in widespread environmental contamination. In addition more toxic and environmentally persistent pesticides are used extensively in developing nations, creating serious acute health problems and local and global environmental impacts (Ecobichon, 2001). Several pesticides were detected in groundwater (Garcia de Llasea and Bernal-Gonzales, 2001; Johnson et al, 2001; and Kadian et al 2008). The amount and type of pesticides in the water of a particular area depends largely on the intensity of production and type of crops being cultivated (Belmonte et al, 2005). Also, the rate at which a pesticide is degraded in both surface and subsurface soils is an important factor in determining the groundwater contamination potential of the pesticide (Di et al, 1998). This implies that most of the applied pesticides find their way as 'residue' in the environment into water and the terrestrial and aquatic food chains where they undergo concentration and exert potential, long term, adverse health effects (Ekstrom et al, 1996; Hirone et al, 2000; and Osman and Al-Rehiyani, 2003). Since the late decades, concern about the contamination of water sources has risen due to the increasing number of pesticides detected. Regulations for drinking water are required in order to limit human risks and environmental pollution. These regulations are well defined in North America, depending on the toxicity level of each compound, and Europe (Directive, 1998) setting at 0.1 ppm of pesticide concentration for a single pesticide compound and 0.5 ppm for the sum of all pesticides in water samples. Chlorpyrifos (O,O-diethyl O-3,5,6-trichloro-2-pyridyl phosphorothioate) is one of the most organophosphorus pesticides that is a widely-used for controlling various insect pests in agricultural and urban settings. Chlorpyrifos enters aquatic systems through spray drift, runoff, erosion, and spills (Racke, 1993). Cypermethrin has been widely used in both indoors and outdoors (Kaufman et al, 1981). It is especially effective towards the control of insect pests in many crops, outdoor mosquito control and as an indoor insecticide (Takahashi et al, 1985). Cypermethrin is commonly found in rivers, sediments, soils, and even foodstuffs (Allan et al, 2005 and Amweg et al, 2005). In recent times the removal of organic harmful pollutants present is investigated by means of a variety of chemical procedures. Advanced oxidation processes (AOPs) which are constituted by the combination of several oxidants, have proven to be very effective in treating a wide variety of organic contaminants. These technologies utilize powerful oxidizing intermediates (mainly OH radicals) to oxidize organic pollutants, leading not only to their destruction, but also, given sufficient conditions, to their complete mineralization. The OH radicals can be generated, for example, by the application of ozone/hydrogen

peroxide, ultraviolet radiation/ozone, ultraviolet radiation/hydrogen peroxide, ozone/electron beam (Gehring et al, 1992;Legrini et al, 1993 andAcero et al 2001). Ozone is a triatomic form of oxygen and is referred to as activated oxygen, allotropic oxygen or pure air. It is an unstable gas and the half-life ozone in distilled water at 20 oxygen, and even more rapidly in impure solution (Hill and Rice 1982), while It has a long half-life in the gaseous state ??Rice, 1986). Ozone has been approved for use as a disinfectant or sanitizer in foods and food processing in the United States for removing residual pollutants such as pesticides and other pollutants which are difficult to get rid of in biological oxidation processes due to its high oxidability, high reaction rate and absence of any secondary pollution. It is considered as a powerful oxidant having electrochemical oxidation potential of 2.0V versus 2.8V for hydroxyl radical. Consequently, oxidation by ozone have usually been used as an effective method for removing residual pollutants such as pesticides and other hazardous chemicals from raw water used for drinking and for wastewater treatment (Lafi and Al-Qodah, 2006). There is no data about the removal of chropyrifos or cypermethrin from water in KSA and the search for means to improve the production of clean water in KSA is always the target of scientists, politicians and businessmen, who seek new techniques to enhance the quality and safety of this product. A wide range of water and terrestrial ecosystems might be contaminated with chlorpyrifos and cypermethrin (EPA, 1997 and Sapozhnikova et al, 2004) which have increased the public concern to establish an efficient, safe, and cost effective method to remove or detoxify chlorpyrifos and cypermethrin residues in contaminated water. Therefore, the present study was carried out to evaluate the effectiveness of ozone at different contact times as a safe method for removal of chropyrifos and cypermethrin in water samples.

## 2 II.

## 3 Materials and Methods

### 4 a) Chemicals b) Experimental Procedure i. Removal of organic compounds residues from groundwater

Groundwater samples were collected from different locations at Al-Qassim region. Water samples were fortified with the organic compounds, chlropyrifos and cypermethrin at two levels for each compound (1 and 2 ppm). Removal of organic compounds from water samples was studied at four different contact times to ozone gas (15, 30, 45 and 60 min) at room temperature were treated with ozone at the previous mentioned periods. Ozone gas (100 ppm at air flow rate of 2.5 L/min with ozone output of 300 mg/hr) produced by Xetin Ozone Air &Water purifier (Model XT 301, Xetin Co. Ltd, Taiwan) was bubbled into 10 liters deionized water in polypropylene cylinder. The duration of dissolved ozone levels were controlled via adjusting the duration of bubbling. A 500 ml volume of water was withdrawn after different time on intervals and kept at analysis.

ii. Extraction Procedure Water samples were extracted according to the procedure of Quintana et al (2001). with slight modification. In brief, A 500 ml volume of water, in which 5 ml of methanol had been added, was passed over the conditioned sorbent (3 ml of ethyl acetate, 3 ml of methanol and 6 ml of water, the sorbent of SPE was never allowed to dry during the conditioning and sample loading steps.) at a flow-rate of 5 ml/min. The sorbent was afterwards dried under vacuum for 20 min. Elution was performed by soaking the cartridge with 2.5 ml of ethyl acetate at a flow-rate of 0.8 ml/min, eluted with a second portion of 2.5 ml of ethyl acetate and collected in a glass vessel containing 0.5 ml of isoctane. The elute was then dried under vacuum to 0.45 ml. The volume was precisely readjusted to 0.5 ml of isoctane and then analyzed by gas chromatography-mass spectroscopy (GC-MS).

iii. Recovery Experiments Pure water samples were spiked with either chlropyrifos or cypermethrin standard solutions in acetone to give the four levels of 0.25, 0.50, 1 and 2 ppm prior to extraction. They were then prepared according to the proposed procedure as described previously and then absolute recoveries were measured. The recovery values were found to be ranged from 98-104 and 92-106% for chlropyrifos and cypermethrin, respectively.

### 5 iv. Determination of detection and quantitation limits

Remediation of Pesticide-Polluted Water Using Ozonation as a Safe Method© 2013 Global Journals Inc. (US)

The limits of detection (LOD) and limit of quantitation (LOQ) were calculated from the signal-tonoise ratios obtained by analyzing unspiked samples ( $n = 10$ ); LOD and LOQ were taken to be the concentrations of pesticide resulting in a signal-to-noise ratio of 3 and 10, respectively. The LOD values were Gas chromatography (Model GC 450, Varian Inc., The Netherlands) with a mass spectrometry (MS 220.41) detector equipped with split/splitless injector with electronic pressure control was employed. A Fused silica CP-Sil 8 CB-LB/MS capillary column (30 m x 0.25 mm i.d) was used in combination with the following oven temperature programme for chlropyrifos: initial temperature 50 °C, 5 °C/min ramp to 160 °C held for 10 min (first step) and from 160 to 250 °C (20 min) at 15 °C (final step) and 90 °C, 5 °C/min ramp to 160 °C held for 10 min (first step) and from 160 to 250 °C (20 min) at 15 °C (final step) for cypermethrin. The injector temperature was 280 °C and mass range from 50-650 amu. The carrier gas (helium, 99.999%) flow rate was set to a constant head pressure of 200 kPa at flow rate of 1.0 ml min<sup>-1</sup> with split ratio of 1: 20 min. The mass spectrometer was operated in electron ionization mode with a transfer line temperature of 280 °C, manifold temperature 40 °C, ion trap temperature 200 °C, ion source

240 °C and selected ion monitoring (SIM) mode. The ion energy for electron impact (EI) was kept at 70 eV. MS Workstation version 6.9.1. was used for data acquisition. For positive identification, both retention time (Rt) and the presence of five fragment ions (z/m ions: 197, 97, 199, 29 and 414 for chlproprifos and 163, 165, 181, 91 and 77 for cypermethrin) were considered. Figures ?? and 2 represent the GC-MS chromatograms for chlproprifos and cypermethrin, respectively.

## 6 III.

## 7 Results and Discussion

The study shows one of the analytical methods that use a solid phase extraction (SPE) to pre-treat the sample and further analysis of the extract by gas chromatograph-mass spectrometer (GC-MS) equipped with electron impact ionization (EI) detector. SPE is particularly suited for the isolation of organic micropollutants from water and has now become the method of choice in order to carry out simultaneously the extraction and concentration of many pesticides and metabolites in aqueous samples (Heberer et al, 1994; Font, 1993 and ??abik et al, 2000). The most widely used sorbents are C8 and C18 chemically bonded to silica, carbon black and polymeric resins ??Sabik et al, 2000).

### 8 a) Removing of chlproprifos and cypermethrin by ozone treatment

The effect of ozone treatment on either chlproprifos or cypermethrin residues for different contact times was investigated. The amount of either chlproprifos or cypermethrin levels was significantly decreased exponentially as the contact time increased in water samples at the two tested levels of the pesticides (1 and 2 ppm) compared to the initial levels, control, (Tables ??-2). The data showed that ozone declined the amount of chlproprifos in water samples following the all intervals of treatment. The removal percentages were 10.5, 96.3, 97.4 and 98.5% in samples spiked with 1ppm of chlproprifos after 15, 30, 45 and 60 min, respectively, while they were 79.6, 93.9, 94.7 and 96.1% in samples spiked with 2 ppm. In case of cypermethrin, the removal percentages were 68.6, 90.5, 97 and 99.2% in water samples spiked with 1ppm following ozone bubbling for 15, 30, 45 and 60 min, respectively, whereas they were 30.5, 50, 94.7, 84.7 and 92% at 2 ppm fortification level in the same order.

### 9 b) Kinetic Studies

A biphasic model was assumed according to Sigma Plot (2011) in order to carry out the statistical study of the of either chlproprifos or cypermethrin removal in water (equation 1).  $C_t = A_0 e^{-k_1 t} + B_0 e^{-k_2 t}$   $t_{1/2} = (2.303 \log 2) / \text{rate constant}(2)$

The data fitting results in case of all ozone treatment using second order kinetic showed that the coefficients of determination (R<sup>2</sup>) were 0.829 and 0.999 for chlproprifos as well as 0.9990 and 0.9510 for cypermethrin when the tested levels of pesticide were 1 and 2 ppm, respectively (Tables ?? and 6). The biphasic model is characterized by a rapid phase (first phase), and a much slower phase (second phase). This is clearly reflected in the half-live values (t<sub>1/2</sub>), where t<sub>1/2</sub> values for chlproprifos were 15.0 and 4 min, and t<sub>1/2</sub> values were 15.1 and 17.96 min, at the spiking levels of 1 and 2 ppm, respectively, following ozone treatment. On the other hand, t<sub>1/2</sub> values of cypermethrin were 8.89 and 21.71 min and t<sub>1/2</sub> values were 9.00 and 21.71 min at the spiking levels of 1 and 2 ppm, respectively, following ozone treatment.

The present findings are in accordance with those of many investigators who reported that the kinetics of pesticide degradation is commonly biphasic with a very rapid degradation rate at the beginning followed by a very slow prolonged dissipation ??Alexander, 1994; Jones et al, 1996; ??igas et al, 2007 and Osman et al, 2009). The relative importance of the phases depends on the availability of the pollutants, hydrophobicity, and affinity for organic matter. So it is recommended to use such simple and non-toxic Year 2013 B related to its water solubility and octanol-water partition coefficient.

One of the health concerns of using oxidants to degrade pesticide is the formation of toxic intermediates. The present study investigated the efficacy of ozone to remove chlproprifos and cypermethrin from water. Ozone was assayed for treatment has a powerful oxidant having electrochemical oxidation potential of 2.0V, and thus, can modify the chemical structure of the selected pesticides creating derived by-products. If these by-products are more toxic than the parent pesticide, such washing treatments should not be utilized to reduce pesticide residue levels in water.

Ozone selectively reacts with compounds containing hetero-atoms such as S, N, O, and Cl via two different pathways, namely direct molecular and indirect radical chain-type reactions (Gottschalk et al, 2000). Thus, pesticides, which usually have some hetero-atoms on the molecules, are often expected to be destroyed by ozonation (Reynolds, 1989). However, as has been found by many researchers, the reactivity of pesticides with ozone varies largely due to their diverse structural features (Reynolds, 1989; David et al, 1991) the characteristics of the wastewater to be treated, i.e., pH, concentration of ozone decomposition initiators, promoters and scavengers in the reacting medium (Glaze et al, 1987).

## IV.

## 10 Conclusions

Water is the basic necessity of life and water contaminated with toxic pesticides is associated with severe effects on the human health. Hence it is pertinent to explore strategies that address this situation of water safety especially for the developing countries where pesticide contamination is widespread due to indiscriminate usage. It is therefore of significance to evaluate simple and effective strategies as such ozone to enhance water safety from harmful pesticides. Due to its high oxidability, high reaction rate and absence of any secondary pollution, ozonolysis technique should be used in the sanitization of water especially in the treatment of pesticides which are difficult to get rid of. Results of this work, provided some basic concepts that can be helpful in water treatment for consumers. Therefore, the present study validated that ozone treatment is safe and promising processes for the removal of pesticides from water under domestic conditions. Results found in the present study must not be extrapolated to other pesticides.

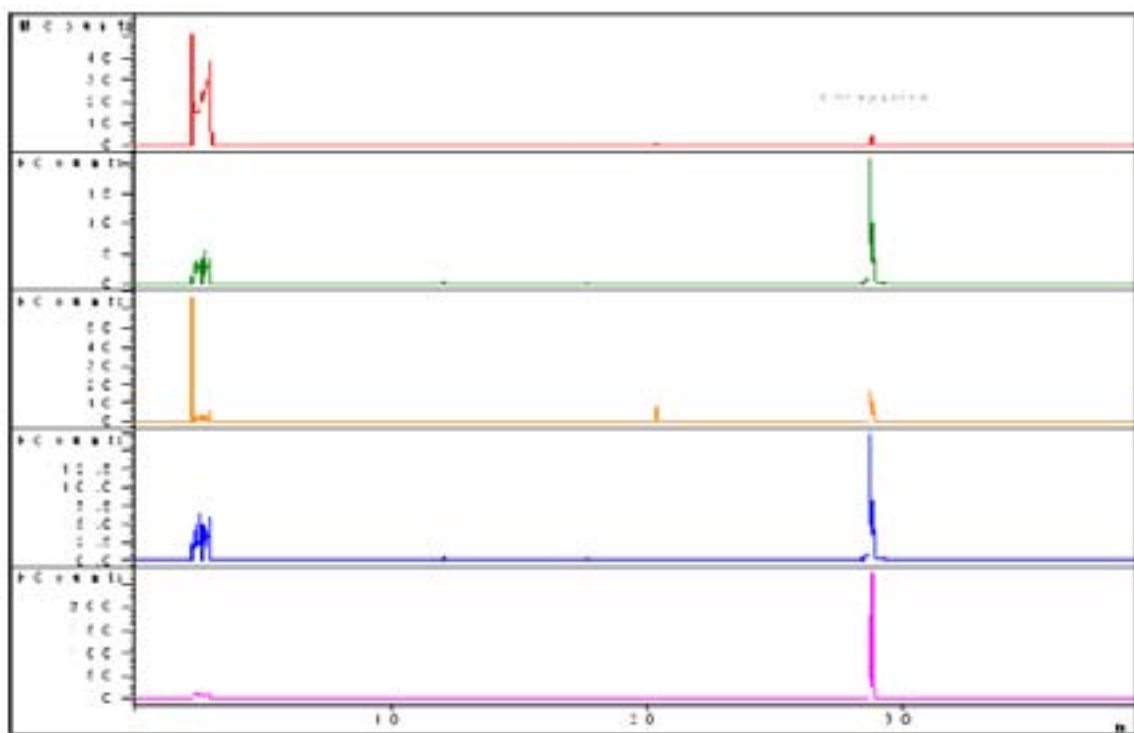


Figure 1:

---

<sup>1</sup>© 2013 Global Journals Inc. (US)

<sup>2</sup>© 2013 Global Journals Inc. (US) 2 20



**Fig (1). GC-MS profiles for chlorpyrifos**

Figure 2: Global



## 1 Acknowledgements

This study was supported by the Engineering and Applied Sciences Research Center, Scientific Research Deanship, Majmaah University under Award No. 11-1432.

28. Rice RG. ??1986)

[Font et al. ()] , G ; J Font , ; J C Manes , Y Molto , Pico . *J. Chromatography* 1993. 642 p. 135.

[Heberer et al. ()] , T Heberer , ; S Butz , H J Stan . *Journal of AOAC International* 1994. 77 p. 1587.

[Ec Directive ()] 98/ 83, *Publicatieblad Europese Gemeenschappen* 330/ 32-330, Ec Directive . 1998. 1998. 54.

[Reynolds et al. ()] 'Aqueous ozonation of pesticides: A Review'. G Reynolds , N Graham , R Perry , R G Rice . *Ozone Science & Engineering* 1989. 11 p. .

[Kadian et al. ()] 'Biodegradation of herbicide (atrazine) in contaminated soil using various bioprocessed materials'. N Kadian , A Gupta , S Satya , R K Mehta , A Malik . *Bioresource Technology* 2008. 99 p. .

[Osman et al. ()] 'Bioremediation of oxamyl in sandy soil using animal manures'. K A Osman , S Al-Rehiyani , Al-Doghairi , M A Salama , AK . *International Biodeterioration & Biodegradation* 2009. 63 p. .

[Lafi and Al-Qodah ()] 'Combined advanced oxidation and biological treatment processes for the removal of pesticides from aqueous solutions'. W K Lafi , Z Al-Qodah . *Journal of Hazardous Materials* 2006. 137 p. .

[Di et al. ()] 'Degradation rates of eight pesticides in surface and subsurface soils under laboratory and field conditions'. H Di , L A G Aylmore , R S Kookana . *Soil Science* 1998. 163 p. .

[Racke ()] *Environmental fate of chlorpyrifos. Reviews of Environmental Contamination & Toxicology*, K D Racke . 1993. 131 p. .

[Hill and Rice (ed.) ()] *Historical background properties and applications*, A G Hill , R G Rice . Rice, R.G. (ed.) 1982. Ann Arbor, MI: Ann Arbor Science Publishers. p. . (Ozone Treatment of Water for Cooling Application)

[Vega et al. ()] 'Monitoring of pesticides in agricultural water and soil samples from Andalusia by liquid chromatography coupled to mass spectrometry'. Belmonte Vega , A Garrido Frenich , A Vidal , JL . *Analytica Chimica Acta* 2005. 538 (1-2) p. .

[Quintana et al. ()] 'Monitoring of pesticides in drinking and related waters in NE Spain with a multiresidue SPE-GC-MS method including an estimation of the uncertainty of the analytical results'. J ; I Quintana , F Martí , Ventura . *Journal of Chromatography A* 2001. 2001. 938 p. .

[Kaufman et al. ()] 'Movement of cypermethrin, decamethrin, permethrin, and their degradation products in soil'. D D.; B A Kaufman , S Russell; C , A J Hellingand , Kayser . *Journal of Agricultural & Food Chemistry* 1981. 29 p. .

[Acero et al. ()] 'MTBE oxidation by conventional ozonation and the combination ozone/hydrogen peroxide: efficiency of the processes and bromate formation'. Juan L Acero , Stefan B Haderlein , C Torsten , Schmidt , J F Marc , Suter , Urs Von Gunten . *Environmental Science & Technology* 2001. 35 p. .

[Jones et al. ()] 'Organic chemicals in contaminated land: analysis, significances and research priorities'. K C Jones , R E Alcock , D L Johnoson , G L Nothcott , K T Semple , P J Woolgar . *Land Contamination & Reclamation* 1996. 3 p. .

[Gottschalk et al. ()] *Ozonation of Water and Waste Water: A Practical Guide to Understanding Ozone and its Application*, C Gottschalk , J A Libra , A Saupe . 2000. Weinheim: Wiley-VCH. p. .

[Johnson et al. ()] 'Penetration of herbicide to groundwater in an unconfined chalk aquifer following normal soil applications'. A C Johnson , T J Besien , A Bhardwaj , D C Dixon , A H Gooddy , Haria , C White . *Journal of Contaminant Hydrology* 2001. 53 p. .

[Chiron et al. ()] 'Pesticide chemical oxidation: state-of-the-art'. S Chiron , A Fernandez-Alba , E Rodriguez , Garcia-Calvo . *Water Research* 2000. 34 p. .

[Ecobichon ()] 'Pesticide use in developing countries'. D J Ecobichon . *Toxicology* 2001. 160 (1-3) p. .

[Legrini et al. ()] 'Photochemical processes for water treatment'. O Legrini , E Oliveros , A M Braun . *Chemical Reviews* 1993. 93 p. .

[García De Llasea and Bernal-González ()] 'Presence of carbamate pesticides in environmental waters from the northwest of Mexico: Determination by liquid chromatography'. M P García De Llasea , M Bernal-González . *Water Research* 2001. 35 p. .

[David et al. ()] 'Rate constants for direct reactions of ozone with several drinking water contaminants'. C David , C Yao , W R Haag . *Water Research* 1991. 25 (7) p. .

[Gehring et al. ()] 'Remediation of ground water polluted with chlorinated ethylenes by ozone electronbeam irradiation treatment'. P; Gehring , Proksch , H Eschweiler , W Szinovatz . *International Journal of Radiation Application and instrumentation. Part A. Applied Radiation and Isotopes* 1992. 45 p. .

- 226 [Epa ()] 'Review of Chlorpyrifos Poisoning Data'. Epa . *EPA* 1997.
- 227 [Osman and Al-Rehiyani ()] 'Risk assessment of pesticide to human and the environment'. K A Osman , S  
228 Al-Rehiyani . *Saudi Journal of Biological Sciences* 2003. 10 p. .
- 229 [Glaze et al. ()] 'The chemistry of water treatment processes involving ozone, hydrogen peroxide and ultraviolet  
230 irradiation'. W H Glaze , J W Kang , D H Chapin . *Ozone Science Engineering* 1987. 9 p. .
- 231 [Amweg et al. ()] 'Use and toxicity of pyrethroid pesticides in the central valley California, USA'. E L Amweg ,  
232 D P Weston , N M Ureda . *Environmental Toxicology & Chemistry* 2005. 24 (4) p. .